

Available online at www.sciencedirect.com



Tetrahedron Letters

Tetrahedron Letters 48 (2007) 1331-1335

## Total synthesis of siomycin A

Tomonori Mori, Shuhei Higashibayashi,<sup>†</sup> Taiji Goto, Mitsunori Kohno, Yukiko Satouchi, Kazuyuki Shinko, Kengo Suzuki, Shunya Suzuki, Hiraku Tohmiya, Kimiko Hashimoto<sup>\*,‡</sup> and Masaya Nakata<sup>\*</sup>

Department of Applied Chemistry, Faculty of Science and Technology, Keio University, 3-14-1 Hiyoshi, Kohoku-ku, Yokohama 223-8522, Japan

Received 4 December 2006; revised 16 December 2006; accepted 21 December 2006 Available online 23 December 2006

Dedicated to Professor Yoshito Kishi on the occasion of his forthcoming 70th birthday

Abstract—We have succeeded in the total synthesis of siomycin A, a representative compound of the thiostrepton family of peptide antibiotics, featuring the one-pot cyclization–elongation of our strategic intermediates and the late-stage formations of the thiazoline and dehydroamino acid moieties. © 2007 Elsevier Ltd. All rights reserved.

Siomycin A was isolated in 1961 from the culture broth of *Streptomyces sioyaensis* by the Shionogi group<sup>1</sup> and is a representative of the thiostrepton family of peptide antibiotics (Fig. 1).<sup>2</sup> The structure of siomycin A was elucidated by chemical degradation studies<sup>3</sup> and NMR spectral studies<sup>4</sup> by comparison with those of thiostrepton. Other structurally-related antibiotics, the thiopeptins, Sch 18640, and Sch 40832, were also isolated.<sup>2</sup> The characteristic structure of this family is the bicvclic skeleton containing dehvdropiperidine, dihvdroquinoline, four thiazoles, thiazoline, dehydroamino acids, and dihydroxyisoleucine. Fascinated by their stunningly complex structural features, we<sup>5</sup> and the Nicolaou  $\operatorname{group}^6$  have aimed to synthesize these antibiotics. Recently, Nicolaou and his co-workers have succeeded in the fascinating total synthesis of thiostrepton.<sup>7</sup> Other efforts have focused on the syntheses of the structurally simpler thiopeptide antibiotics: for example, the micrococcins,<sup>8</sup> promothiocin A,<sup>9</sup> amythiamicin D,<sup>10</sup>

0040-4039/\$ - see front matter @ 2007 Elsevier Ltd. All rights reserved. doi:10.1016/j.tetlet.2006.12.121



Figure 1. Structure of siomycin A.

GE2270A,<sup>11</sup> and GE2270T.<sup>11</sup> We now report the total synthesis of siomycin A by the coupling (① in Fig. 1) of our strategic intermediates, the cyclic core portion  $1^{5e}$  (A-ring, Fig. 1 and Scheme 1) and the pentapeptide portion  $2^{5c}$  (Scheme 1), followed by cyclization (② in Fig. 1, B-ring) of the resulting coupling product and elongation (③ in Fig. 1) of the side chain 3 (Scheme 2) onto the cyclization product.<sup>12</sup> We anticipated that this

*Keywords*: Siomycin A; One-pot cyclization-elongation; Thiazoline; Dehydroamino acid.

<sup>\*</sup> Corresponding authors. Tel.: +81 75 595 4673; fax: +81 75 595 4763 (K.H.); tel.: +81 45 566 1577; fax: +81 45 566 1551 (M.N.); e-mail addresses: kimikoh@mb.kyoto-phu.ac.jp; msynktxa@applc.keio. ac.jp

<sup>&</sup>lt;sup>†</sup>Present address: Research Center for Molecular-Scale Nanoscience, Institute for Molecular Science, Okazaki 444-8787, Japan.

<sup>&</sup>lt;sup>‡</sup>Present address: Kyoto Pharmaceutical University, 1 Shichono-cho, Misasagi, Yamashina-ku, Kyoto 607-8412, Japan.



Scheme 1. Synthesis of the intermediate 11. Boc = tert-butoxycarbonyl, Bpoc = 1-methyl-1-(4-biphenyl)ethoxycarbonyl, Fm = 9-fluorenylmethyl, TBS = tert-butyldimethylsilyl, TMSE = 2-(trimethylsilyl)ethyl, HATU = O-(7-azabenzotriazol-1-yl)-1,1,3,3-tetramethyluronium hexafluorophosphate, TES = triethylsilyl, Teoc = 2-(trimethylsilyl)ethoxycarbonyl, DAST = (diethylamino)sulfur trifluoride.

cyclization–elongation step would be realized in the stepwise manner after the selective deprotection of one of the two TMSE esters, or more conveniently, in the one-pot operation via the regioselective cyclization–elongation of the dicarboxylic acid secured by the simultaneous deprotection of the two TMSE esters. Other challenging tasks included the well-timed constructions of the easily racemizing thiazoline ring and the fastidious dehydroamino acids including four dehydroalanines and the trisubstituted (Z)-olefin next to the thiazoline ring.

We have already reported the synthesis of the siomycin cyclic core portion  $1^{5e}$  by the coupling of the dehydropiperidine segment 4,<sup>5a,e</sup> the dihydroquinoline segment 5,<sup>5b,d</sup> and the masked dehydroalanine (i.e.,  $\beta$ -phenylselenoalanine) segment  $6^{5e}$  (Scheme 1). For the coupling partner of 1, we selected  $2^{5c}$  which has the hydroxymethylthioamide function as the precursor of the thiazoline ring (Scheme 1). The Boc group in 1 was first deprotected with 4 M HCl/dioxane to afford 7 along with a small amount of 8. This crude mixture was coupled with 2 (1.0 equiv) using HATU<sup>13</sup> (1.0 equiv) and (*i*-Pr)<sub>2</sub>NEt (5.0 equiv) in CH<sub>2</sub>Cl<sub>2</sub>, giving 9 and its TBS ether 10 in 60% and 8% yields, respectively.

We first attempted selectively deprotecting one of the two TMSE esters in 9. It was anticipated that ZnCl<sub>2</sub>,

which had been used as a deprotection reagent of the Teoc group,<sup>14</sup> would be suitable for the deprotection of the TMSE ester, and additionally, the simultaneous deprotection of the Teoc and acetonide groups would occur. However, under the restricted conditions (ZnCl<sub>2</sub> (100 equiv)/ether, nitromethane, rt, 24 h), we could not realize this requirement; the dicarboxylic acid and a mixture of the monocarboxylic acids were nonselectively obtained, although the Teoc and acetonide groups were smoothly cleaved.<sup>5c</sup>

We then turned our attention to the regioselective cyclization–elongation of the dicarboxylic acid. Prior to this, **9** was first treated with DAST<sup>5c,15</sup> (1.6 equiv) in CH<sub>2</sub>Cl<sub>2</sub> to give thiazoline **11** in 87% yield (Scheme 1). Deprotection of the three kinds of protecting groups (Teoc, acetonide, and TMSE) in **11** was cleanly realized with ZnCl<sub>2</sub><sup>14</sup> (100 equiv)/ether in nitromethane (rt, 48 h), giving the cyclization–elongation precursor **12** (Scheme 2). The side chain **3**<sup>7c</sup> was prepared from the known β-phenylselenoalanine **13**<sup>5e,7c</sup> by the following four-step sequence: (1) amidation, (2) deprotection of the Boc group, (3) condensation of another **13**, and (4) deprotection of the Boc group. The one-pot operation was carried out under the conditions including, as the cyclization step, EDC–HOAt–(*i*-Pr)<sub>2</sub>NEt–DMF (EDC = (1-[3-(dimethylamino)propyl]-3-ethylcarbodiimide hydro-



Scheme 2. Completion of the total synthesis of siomycin A. Reagents and conditions: (a) 1 M ZnCl<sub>2</sub>/ether, MeNO<sub>2</sub>, rt, 48 h; (b) ClCO<sub>2</sub>Et, Et<sub>3</sub>N, THF, -15 °C, 10 min, then 28% NH<sub>4</sub>OH, -15 °C to rt, 2 h; (c) TFA-CH<sub>2</sub>Cl<sub>2</sub> (1:1), rt, 1.5 h; (d) 13, CIP, HOAt, (*i*-Pr)<sub>2</sub>NEt, DMF, rt, 2 h; (e) TFA-CH<sub>2</sub>Cl<sub>2</sub> (1:1), rt, 2 h, 68% (4 steps); (f) HATU, (*i*-Pr)<sub>2</sub>NEt, DMF-CH<sub>2</sub>Cl<sub>2</sub> (1:4), 0 °C, 3 h, then 3, rt, 24 h; (g) HF·pyridine-THF (1:4), rt, 20 h; (h) 4 M TBHP/CH<sub>2</sub>Cl<sub>2</sub>, TFE-CH<sub>2</sub>Cl<sub>2</sub> (1:5), rt, 1 h, 7% from 11. TFA = trifluoroacetic acid, CIP = 2-chloro-1,3-dimethylimidazolidium hexafluoro-phosphate, HOAt = 1-hydroxy-7-azabenzotriazole, TBHP = *tert*-butyl hydroperoxide, TFE = 2,2,2-trifluoroethanol.

chloride) and HATU<sup>13</sup>-(*i*-Pr)<sub>2</sub>NEt-solvent (e.g., DMF, THF, dioxane, DMF-CH<sub>2</sub>Cl<sub>2</sub> (1:4), DMF-CH<sub>3</sub>CN (1:4)), followed by the subsequent elongation with 3. Among them, the best conditions were HATU (5.0 equiv) and (*i*-Pr)<sub>2</sub>NEt (5.0 equiv) in DMF-CH<sub>2</sub>Cl<sub>2</sub> (1:4, 1 mM for 12) at 0 °C for 3 h followed by the addition of 3 (5.0 equiv, rt, 24 h), affording the crude products including 14 after removing the excess 3 by Sephadex LH-20 (CHCl<sub>3</sub>). Although the structure of 14 could not be confirmed at this stage, we proceeded to the two-step transformation into siomycin A. These crude products were treated with HF pyridine-THF  $(1:4)^{7d}$  to afford the crude products including 15, which were finally subjected to the oxidative elimination (4 M TBHP/CH<sub>2</sub>Cl<sub>2</sub>, TFE–CH<sub>2</sub>Cl<sub>2</sub> (1:5), rt, 1 h),<sup>5c,e,6c,7b,d</sup> giving siomycin A in 7% yield from 11. The synthetic siomycin A was identical with the natural siomycin A based on the <sup>1</sup>H NMR, <sup>13</sup>C NMR, IR, and MS spectra, TLC, and optical rotation (see Supplementary data). It

is noteworthy that the final two-step operation could not be reversed, in contrast to Nicolaou's thiostrepton synthesis,7b,d because it was found that siomycin A gradually changed into siomycin B,<sup>16</sup> which is the side chain degradation product of siomycin A, under the HF pyridine-THF (1:4) conditions (rt, during 24 h). In addition, when the model pentapeptide  $16^{5c}$  was subjected to the HF pyridine-THF (1:4) conditions (rt, 4 h), the Z-olefin 17 was obtained in 70% yield as the sole product (Scheme 3). The stereochemistry of 17 was confirmed by NOE analysis of the <sup>1</sup>H NMR spectrum, and additionally, by the transformation (TESOTf, 2,6-lutidine) into 18 which was identical to the sample<sup>5c</sup> derived from 16 by syn elimination using TBHP. These facts mean that the dehydroselenation next to the thiazoline C2-position affords the thermodynamically stable Z-olefin and account for the Z-selectivity from 14 to 15. Unfortunately, during the cyclization-elongation step, an almost equal amount of the product was obtained whose struc-



Scheme 3. Dehydroselenation of the model pentapeptide 16. Tf = trifluoromethanesulfonyl.

ture was tentatively assigned, after HF·pyridine and TBHP treatment and subsequent separation from siomycin A, to the regioisomeric cyclization–elongation product based on the <sup>1</sup>H NMR and MS spectra (see Supplementary data).

In summary, we have succeeded in the total synthesis of siomycin A, a representative compound of the thiostrepton family of peptide antibiotics, by the coupling of the three intermediates, 1, 2, and 3. The drawback in the cyclization-elongation step will be improved once the dehydropiperidine segment having the differentiated protecting groups instead of the bis-TMSE ester in 4 is secured, which is now in progress in our laboratories.

## Acknowledgments

We thank Dr. Kazuyuki Minagawa (Shionogi & Co., Ltd.) for kindly providing natural siomycin A. We are indebted to Mr. Minoru Ogata and Mr. Yasushi Ono (Japan Tobacco Inc.) for the measurement of the HSQC and HMBC spectra of siomycin A. This research was partially supported by a Grant-in-Aid for the 21st Century COE program 'KLCC' (T.M.), for Scientific Research on Priority Areas (A) 'Exploitation of Multi-Element Cyclic Molecules' (M.N.), and on Priority Areas 17035076 and 18032067 (M.N.) from the Ministry of Education, Culture, Sports, Science and Technology, Japan (MEXT).

## Supplementary data

Spectral data of natural and synthetic siomycin A and the regioisomeric cyclization–elongation product. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.tetlet. 2006.12.121.

## **References and notes**

- Nishimura, H.; Okamoto, S.; Mayama, M.; Ohtsuka, H.; Nakajima, K.; Tawara, K.; Shimohira, M.; Shimaoka, N. J. Antibiot. Ser. A 1961, 14, 255–263.
- For a review, see: Bagley, M. C.; Dale, J. W.; Merritt, E. A.; Xiong, X. Chem. Rev. 2005, 105, 685–714.
- (a) Ebata, M.; Miyazaki, K.; Otsuka, H. J. Antibiot. 1969, 22, 423–433; (b) Ebata, M.; Miyazaki, K.; Otsuka, H. J. Antibiot. 1969, 22, 434–441.
- (a) Tori, K.; Tokura, K.; Yoshimura, Y.; Okabe, K.; Otsuka, H.; Inagaki, F.; Miyazawa, T. J. Antibiot. 1979, 32, 1072–1077; (b) Tori, K.; Tokura, K.; Yoshimura, Y.; Terui, Y.; Okabe, K.; Otsuka, H.; Matsushita, K.; Inagaki, F.; Miyazawa, T. J. Antibiot. 1981, 34, 124–129; (c) Clayden, N. J.; Inagaki, F.; Williams, R. J. P.; Morris, G. A.; Tori, K.; Tokura, K.; Miyazawa, T. Eur. J. Biochem. 1982, 123, 127–131.
- (a) Higashibayashi, S.; Hashimoto, K.; Nakata, M. *Tetrahedron Lett.* 2002, 43, 105–110; (b) Higashibayashi, S.; Mori, T.; Shinko, K.; Hashimoto, K.; Nakata, M. *Heterocycles* 2002, 57, 111–122; (c) Higashibayashi, S.; Kohno, M.; Goto, T.; Suzuki, K.; Mori, T.; Hashimoto, K.; Nakata, M. *Tetrahedron Lett.* 2004, 45, 3707–3712; (d) Mori, T.; Satouchi, Y.; Tohmiya, H.; Higashibayashi, S.; Hashimoto, K.; Nakata, M. *Tetrahedron Lett.* 2005, 46, 6417–6422; (e) Mori, T.; Tohmiya, H.; Satouchi, Y.; Higashibayashi, S.; Hashimoto, K.; Nakata, M. *Tetrahedron Lett.* 2005, 46, 6423–6427.
- (a) Nicolaou, K. C.; Safina, B. S.; Funke, C.; Zak, M.; Zécri, F. J. Angew. Chem., Int. Ed. 2002, 41, 1937–1940;
   (b) Nicolaou, K. C.; Nevalainen, M.; Safina, B. S.; Zak, M.; Bulat, S. Angew. Chem., Int. Ed. 2002, 41, 1941–1945;
   (c) Nicolaou, K. C.; Nevalainen, M.; Zak, M.; Bulat, S.; Bella, M.; Safina, B. S. Angew. Chem., Int. Ed. 2003, 42, 3418–3424.
- (a) Nicolaou, K. C.; Safina, B. S.; Zak, M.; Estrada, A. A.; Lee, S. H. Angew. Chem., Int. Ed. 2004, 43, 5087–5092; (b) Nicolaou, K. C.; Zak, M.; Safina, B. S.; Lee, S. H.; Estrada, A. A. Angew. Chem., Int. Ed. 2004, 43, 5092– 5097; (c) Nicolaou, K. C.; Safina, B. S.; Zak, M.; Lee, S. H.; Nevalainen, M.; Bella, M.; Estrada, A. A.; Funke, C.; Zécri, F. J.; Bulat, S. J. Am. Chem. Soc. 2005, 127, 11159– 11175; (d) Nicolaou, K. C.; Zak, M.; Safina, B. S.; Estrada, A. A.; Lee, S. H.; Nevalainen, M. J. Am. Chem. Soc. 2005, 127, 11176–11183.
- (a) Kelly, T. R.; Jagoe, C. T.; Gu, Z. Tetrahedron Lett. 1991, 32, 4263–4266; (b) Nakamura, Y.; Shin, C.-g.; Umemura, K.; Yoshimura, J. Chem. Lett. 1992, 1005– 1008; (c) Okumura, K.; Shigekuni, M.; Nakamura, Y.; Shin, C.-g. Chem. Lett. 1996, 1025–1026; (d) Ciufolini, M. A.; Shen, Y. C. J. Org. Chem. 1997, 62, 3804–3805; (e) Shin, C.-g.; Okumura, K.; Shigekuni, M.; Nakamura, Y. Chem. Lett. 1998, 139–140; (f) Okumura, K.; Ito, A.;

Yoshioka, D.; Shin, C.-g. *Heterocycles* **1998**, *48*, 1319–1324; (g) Okumura, K.; Nakamura, Y.; Shin, C.-g. *Bull. Chem. Soc. Jpn.* **1999**, *72*, 1561–1569; (h) Okumura, K.; Suzuki, T.; Nakamura, Y.; Shin, C.-g. *Bull. Chem. Soc. Jpn.* **1999**, *72*, 2483–2490; (i) Ciufolini, M. A.; Shen, Y.-C. Org. Lett. **1999**, *1*, 1843–1846; (j) Fenet, B.; Pierre, F.; Cundliffe, E.; Ciufolini, M. A. Tetrahedron Lett. **2002**, *43*, 2367–2370; (k) Moody, C. J.; Hughes, R. A.; Thompson, S. P.; Alcaraz, L. *Chem. Commun.* **2002**, 1760–1761.

- Total synthesis of promothiocin A: (a) Moody, C. J.; Bagley, M. C. Synlett **1998**, 361–362; (b) Moody, C. J.; Bagley, M. C. Chem. Commun. **1998**, 2049–2050; (c) Bagley, M. C.; Bashford, K. E.; Hesketh, C. L.; Moody, C. J. J. Am. Chem. Soc. **2000**, 122, 3301– 3313.
- Total synthesis of amythiamicin D: (a) Hughes, R. A.; Thompson, S. P.; Alcaraz, L.; Moody, C. J. Chem. Commun. 2004, 946–948; (b) Hughes, R. A.; Thompson,

S. P.; Alcaraz, L.; Moody, C. J. J. Am. Chem. Soc. 2005, 127, 15644–15651.

- Total syntheses of GE2270A and GE2270T: Nicolaou, K. C.; Zou, B.; Dethe, D. H.; Li, D. B.; Chen, D. Y.-K. Angew. Chem., Int. Ed. 2006, 45, 7786–7792.
- 12. In the total synthesis of thiostrepton, Nicolaou et al.<sup>7</sup> chose the first B-ring cyclization and the subsequent A-ring cyclization. They encountered the low-yielding, B-ring cyclization sequence.
- Carpino, L. A. J. Am. Chem. Soc. 1993, 115, 4397– 4398.
- (a) Gioeli, C.; Balgobin, N.; Josephson, S.; Chattopadhyaya, J. B. *Tetrahedron Lett.* **1981**, *22*, 969–972;
  (b) Björkman, S.; Chattopadhyaya, J. *Chem. Scripta* **1982**, *20*, 201–202.
- 15. Lafargue, P.; Guenot, P.; Lellouche, J.-P. Synlett 1995, 171–172.
- Ebata, M.; Miyazaki, K.; Otsuka, H. J. Antibiot. 1969, 22, 364–368.